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| 1 | Деген, Камышный Экспрессия цитоплазматических NOD-2 и RIG-I рецепторов врожденного иммунитета в кишечнике крыс при экспериментальном сахарном диабете // Российский иммунологический журнал.-2014.-№3.-С.525-528 | Degen A., Kamyshny A. Expression of cytoplasmic nod-2 and rig-i receptors of innate immunity in intestine of rats in experimental diabetes mellitus. Russian Journal of Immunology, 2014, no. 3, pp. 525-528 | <https://e.lanbook.com/journal/issue/301555> |
| 2 | Alyanakian M.A., Grela F., Aumeunier A., Chiavaroli C., Gouarin C., Bardel E., Normier G., Chatenoud L., Thieblemont N., Bach J.F. Transforming growth factor-beta and natural killer T-cells are involved in the protective effect of a bacterial extract on type 1 diabetes. Diabetes., 2006, Vol. 55, no. 1, pp. 179-185. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Alyanakian+M.A.%2C+Grela+F.%2C+Aumeunier+A.%2C+Chiavaroli+C.%2C+Gouarin+C.%2C+et+al.+Transforming+growth+factor-beta+and+natural+killer+T-cells+are+involved+in+the+protective+effect+of+a+bacterial+extract+on+type+1+diabetes>. |
| 3 | Biswas A., Banerjee P., Biswas T. Porin of Shigella dysenteriae directly promotes toll-like receptor 2-mediated CD4+ T cell survival and effector function. Mol. Immunol., 2009, Vol. 46, no. 15, pp. 3076-3085. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Biswas+A.%2C+Banerjee+P.%2C+Biswas+T.+Porin+of+Shigella+dysenteriae+directly+promotes+toll-like+receptor+2-mediated+CD4%2B+T+cell+survival+and+effector+function>. |
| 4 | D’Addio F., Fiorina P. Type 1 diabetes and dysfunctional intestinal homeostasis. Trends in Endocrinology & Metabolism, 2016, Vol. 27, no. 7 | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=D%E2%80%99Addio+F.%2C+Fiorina+P.+Type+1+diabetes+and+dysfunctional+intestinal+homeostasis>. |
| 5 | Degen A., Kamyshny A. Distribution characteristics of RIG-I receptors of innate immunity in experimental diabetes mellitus and administration of nonspecific blockers of TNF-α. J Immunol Clin Microbiol., 2018, Vol. 3, no. 3, pp.50-59 | ‑ | <http://dergipark.gov.tr/jicm/issue/38834> |
| 6 | Devaraj S., Dasu M.R., Rockwood J., Winter W., Griffen S.C., Jialal I. Increased toll-like receptor (TLR) 2 and TLR4 expression in monocytes from patients with type 1 diabetes: further evidence of a proinflammatory state. J Clin Endocrinol Metab., 2008, no. 93, pp. 578–583. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Devaraj+S.%2C+Dasu+M.R.%2C+Rockwood+J.%2C+Winter+W.%2C+Griffen+S.C.%2C+Jialal+I.+Increased+toll-like+receptor+(TLR)+2+and+TLR4+expression+in+monocytes+from+patients+with+type+1+diabetes%3A+further+evidence+of+a+proinflammatory+state>. |
| 7 | Dong C. TH17 cells in development: an updated view of their molecular identity and genetic programming. Nat. Rev. Immunol., 2008, no. 8, pp. 337–348. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Dong+C.+TH17+cells+in+development%3A+an+updated+view+of+their+molecular+identity+and+genetic+programming>. |
| 8 | Faustman D.L. TNF, TNF inducers, and TNFR2 agonists: A new path to type 1 diabetes treatment. Diabetes Metab Res Rev., 2018, Vol. 34, no. 1. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/28843039> |
| 9 | Flaherty S., Reynolds J.M. TLR Function in Murine CD4(+) T Lymphocytes and Their Role in Inflammation. Methods Mol Biol., 2016, no. 1390, pp. 215-227 | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/26803632> |
| 10 | Fulford T.S., Ellis D., Gerondakis S. Understanding the Roles of the NF-κB Pathway in Regulatory T Cell Development, Differentiation and Function. Prog Mol Biol Transl Sci., 2015; no. 136, pp. 57-67. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/26615092> |
| 11 | Hase K., Kawano K., Nochi T., Pontes G.S., Fukuda S., Ebisawa M., Kadokura K., Tobe T., Fujimura Y., Kawano S., Yabashi A., Waguri S., Nakato G., Kimura S., Murakami T., Iimura M., Hamura K., Fukuoka S., Lowe A.W., Itoh K., Kiyono H., Ohno H. Uptake through glycoprotein 2 of FimH1 bacteria by M cells initiates mucosal immune response. Nature., 2009, Vol. 462, no 7270, pp. 226-230 | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Uptake+through+glycoprotein+2+of+FimH1+bacteria+by+M+cells+initiates+mucosal+immune+response>. |
| 12 | Imanishi T., Hara H., Suzuki S., Suzuki N., Akira S., Saito T. Cutting edge: TLR2 directly triggers Th1 effector functions. J. Immunol., 2007, no. 178, pp. 6715–6719. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Cutting+edge%3A+TLR2+directly+triggers+Th1+effector+functions>. |
| 13 | Jialal I., Yun J.M., Bremer A., Devaraj S. Demonstration of Increased TLR2 and TLR4 Expression in Monocytes of Type 1 Diabetic Patients with Microvascular Complications. Metabolism, 2011, Vol 60, no. 2, pp. 256–259. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Jialal+I.%2C+Yun+J.M.%2C+Bremer+A.%2C+Devaraj+S.+Demonstration+of+Increased+TLR2+and+TLR4+Expression+in+Monocytes+of+Type+1+Diabetic+Patients+with+Microvascular+Complications>. |
| 14 | Kanaya T., Hase K., Takahashi D., Fukuda S., Hoshino K., Sasaki I., Hemmi H., Knoop K.A., Kumar N., Sato M., Katsuno T., Yokosuka O., Toyooka K., Nakai K., Sakamoto A., Kitahara Y., Jinnohara T., McSorley S.J., Kaisho T., Williams I.R., Ohno H. The Ets transcription factor Spi-B is essential for the differentiation of intestinal microfold cells. Nat Immunol., 2012, Vol. 13, no. 8, pp. 729–736. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/22706340> |
| 15 | Knoop K.A., Kumar N., Butler B.R., Sakthivel S.K., Taylor R.T., Nochi T., Akiba H., Yagita H., Kiyono H., Williams I.R. RANKL is necessary and sufficient to initiate development of antigen-sampling M cells in the intestinal epithelium. J Immunol., 2009, Vol. 183,no. 9, pp. 5738–5747. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Knoop+K.A.+et+al.+RANKL+is+necessary+and+sufficient+to+initiate+development+of+antigen-sampling+M+cells+in+the+intestinal+epithelium>. |
| 16 | Koulmanda M., Bhasin M., Awdeh Z., Qipo A., Fan Z., Hanidziar D., Putheti P., Shi H., Csizuadia E., Libermann T.A., Strom T.B. The role of TNF-α in mice with type 1- and 2- diabetes. PLoS One., 2012, Vol. 7, no. 5, pp. 332-354. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/22606220> |
| 17 | Liang L., Beshay E., Prud'homme G.J.The phosphodiesterase inhibitors pentoxifylline and rolipram prevent diabetes in NOD mice. Diabetes., 1998, Vol. 47, no. 4, pp. 570-575. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/9568689> |
| 18 | Maloy K.J., Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. Nature, 2011, no. 474, pp. 298–306. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/21677746> |
| 19 | Nakamura Y., Kimura S., Hase K. M cell-dependent antigen uptake on follicle-associated epithelium for mucosal immune surveillance. Inflamm Regen., 2018, no. 38, pp. 1-5. | ‑ | <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6120081/> |
| 20 | Needell J.C., Zipris D. The Role of the Intestinal Microbiome in Type 1 Diabetes Pathogenesis. Curr Diab Rep., 2016, Vol. 16, no. 10, pp 8-9. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/27523648> |
| 21 | Ohno H. Intestinal M cells. J Biochem., 2016, Vol. 159, no. 2, pp. 151-160. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/26634447> |
| 22 | Opazo M.C., Ortega-Rocha E.M., Coronado-Arrázola I., Bonifaz L.C., Boudin H., Neunlist M., Bueno S.M., Kalergis A.M., Riedel C.A.. Intestinal Microbiota Influences Non-intestinal Related Autoimmune Diseases. Front Microbiol., 2018, Vol. 9. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/29593681> |
| 23 | Qiao Y.C., Chen Y.L., Pan Y.H., Tian F., Xu Y., Zhang X.X., Zhao H.L. The change of serum tumor necrosis factor alpha in patients with type 1 diabetes mellitus: A systematic review and meta-analysis. PLoS One., 2017, Vol. 12, no. 4. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/28426801> |
| 24 | Ramakrishnan P., Yui M.A., Tomalka J.A., Majumdar D., Parameswaran R., Baltimore D. Deficiency of Nuclear Factor-κB c-Rel Accelerates the Development of Autoimmune Diabetes in NOD Mice. Diabetes., 2016, Vol. 65, no. 8, pp. 2367-2379. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/27217485> |
| 25 | Sato S., Kaneto S., Shibata N., Takahashi Y., Okura H., Yuki Y., Kunisawa J., Kiyono H. Transcription factor Spi-B-dependent and -independent pathways for the development of Peyer's patch M cells. Mucosal Immunol., 2013, Vol. 6, no. 4, pp.838-46. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/23212199> |
| 26 | Taylor R.T., Patel S.R., Lin E., Butler B.R., Lake J.G., Newberry R.D., Williams I.R. Lymphotoxin-independent expression of TNF-related activation-induced cytokine by stromal cells in cryptopatches, isolated lymphoid follicles, and Peyer’s patches. J Immunol., 2007, Vol. 178, no. 9, pp. 5659–5667.  | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Lymphotoxin-independent+expression+of+TNF-related+activation-induced+cytokine+by+stromal+cells+in+cryptopatches%2C+isolated+lymphoid+follicles%2C+and+Peyer%E2%80%99s+patches>. |
| 27 | Vaarala O. Human intestinal microbiota and type 1 diabetes. Curr Diab Rep., 2013, Vol. 13,no. 5, pp. 601–607. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/23934614> |
| 28 | Visser J., Groen H., Klatter F., Rozing J. Timing of pentoxifylline treatment determines its protective effect on diabetes development in the Bio Breeding rat. Eur J Pharmacol., 2002, Vol. 445, no. 1-2, pp. 133-140. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Visser+J.%2C+Groen+H.%2C+Klatter+F.+Timing+of+pentoxifylline+treatment+determines+its+protective+effect+on+diabetes+development+in+the+Bio+Breeding+rat>. |
| 29 | Wood M.B., Rios D., Williams I.R..TNF-α augments RANKL-dependent intestinal M cell differentiation in enteroid cultures. Am J Physiol Cell Physiol., 2016, Vol. 311, no. 3, pp. 498-507. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/27413168> |
| 30 | Zanin-Zhorov A., Tal-Lapidot G., Cahalon L., Cohen-Sfady M., Pevsner-Fischer M., Lider O., Cohen I.R. Cutting edge: T cells respond to lipopolysaccharide innately via TLR4 signaling. J. Immunol., 2007, Vol. 179, no. 1, pp. 41–44. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/?term=Cutting+edge%3A+T+cells+respond+to+lipopolysaccharide+innately+via+TLR4+signaling>. |
| 31 | Zhang H., Bi J., Yi H., Fan T., Ruan Q., Cai L., Chen Y.H., Wan X. Silencing c-Rel in macrophages dampens Th1 and Th17 immune responses and alleviates experimental autoimmune encephalomyelitis in mice. Immunol Cell Biol., 2017, Vol. 95, no. 7, pp. 593-600. | ‑ | <https://www.ncbi.nlm.nih.gov/pubmed/28202908> |