**Названия рисунков**

Figure 1. Effect of MVs Derived From Cells of the NK-92 Cell Line on Migratory Activity and Proliferation of ECs of the EA.Hy926 Cell Line. (A) Number of cells that migrated to the disrupted monolayer zone; (B) Residual area after migration of cells to the disrupted monolayer zone; (C) Number of cells that proliferated. \*\* - p<0.01; \*\*\* - p<0.001 – difference compared with cells incubated without MVs (2.5% FCS); ## - p<0.01, ### - p<0.001 – difference compared with a lower concentration under the same conditions.

Figure 2. Expression of Surface Receptors by ECs After Culturing With (ECs+MVs) and Without (MVs) MVs Derived From Cells of the NK-92 Cell Line [median (upper quartile, lower quartile)]. Data are presented as the relative number of ECs expressing (a) CD31, CD34, (c) VEGFR1, CD119, and (e) CD105, CD54, and on the intensity of (b) CD31, CD34, (d) VEGFR1, CD119, and (f) CD105, CD54 expression by ECs. Significance of differences compared with unstimulated ECs: \*\* - p<0.01; \*\*\* - p<0.001.

Figure 3. ECs carrying the CD45 receptor after culturing with (ECs+MVs) and without (MVs) MVs. Data on the relative number of ECs with the CD45+ (a) phenotype and on the intensity of fluorescence of ECs with the CD45+ (b) phenotype are presented. Significance of differences compared with unstimulated ECs: \*\* - p<0.01.

Figure 4. ECs of the EA.Hy926 cell line cultured with MVs derived from cells of the NK-92 cell line leads to granzyme B transfer from MVs to ECs. (A) Representative immunoblot for granzyme B in cells of the NK-92 cell line (1) and in their MVs (2). (B) Representative immunoblot for granzyme B in unstimulated cells of the EA.Hy926 cell line (1) and after culture with MVs derived from cells of the NK-92 cell line (2).

Figure 5. Caspase-8 Activation (Defined as the Ratio of Caspase-8 Fragment Intensity to Procaspase-8) in ECs of the EA.Hy926 Cell Line After Culture with MVs Derived From Cells of the NK-92 Cell Line. (A) No difference in caspase-8 activation or procaspase-8 cleavage up to the p18 fragment of active caspase-8 was observed between the ECs after culture with MVs and unstimulated ECs. (B) EC treatment with MVs resulted in activation of caspase-8 and cleavage of procaspase-8 up to the p43/p41 fragment compared with unstimulated cells. (C) Quantitative assessment of procaspase-8, p43/p41 fragment and p18 fragment levels in ECs after culture with MVs. (D) Representative immunoblot demonstrating the effect of MVs derived from cells of the NK-92 cell line on the intensity of caspase-8 fragments (p43/p41 and p18) and procaspase-8 in ECs (1 – unstimulated ECs, 2 – ECs after culture with MVs). \*\*\* - p<0.001 – difference between the indicator in ECs after their culture with MVs and the indicator in unstimulated ECs.

Figure 6. Caspase-3 Activation (Defined as the Ratio of Active Caspase-3 to Procaspase-3 Fragment Intensity) in ECs of the EA.Hy926 Cell Line After Culture with MVs Derived From Cells of the NK-92 Cell Line. (A) EC treatment with MVs results in activation of caspase-3 as compared with unstimulated ECs. (B) Quantitative assessment of procaspase-3 and active caspase-3 fragment levels in ECs after culture with MVs. (C) Representative immunoblot demonstrating the effect of MVs on the intensity of cleaved caspase-3 fragments and on procaspase-3 in ECs (1 – unstimulated ECs, 2 – ECs after culture with MVs). \* - p<0.05; \*\* - p<0.01; \*\*\* - p<0.001 – difference between the indicator in ECs after culture with MVs and the indicator in unstimulated ECs.

Figure 7. Effect of MVs Derived from Cells of the NK-92 Cell Line on Caspase-3 and Caspase-9 Activity in ECs of the EA.Hy926 Cell Line. Caspase-3 activity was assessed by spectrophotometric determination of p-nitrianiline (pNA) cleaved by caspase-3 and caspase-9 from the caspase-specific substrates, DEVD-pNA and Ac-LEHD-pNA, respectively. Activity was measured in μm of released pNA/min/mg of total cell lysate protein. \* - p<0.05; \*\*\* - p<0.001 – difference between the indicator in ECs after culture with MVs and the indicator in unstimulated ECs.

Supplementary Figure 1. Migration of Cells of the EA.Hy926 Cell Line. Stained with crystal violet, x100. A. Initial width of the disrupted monolayer (migration surface) line. В. Migration after incubation in medium containing 2.5% FCS for 24 hours. C. Migration after incubation in medium containing 10% FCS for 24 hours.

Supplementary Figure 2. Phenotypic Characteristics of Unstimulated ECs of the EA.Hy926 Cell Line. 1) Distribution of ECs in a FSC/SSC histogram; 2) Isotypic control; 3) Distribution of ECs treated with antibodies to VEGFR1 (a), VEGFR2 (b), CD34 (c), CD105 (d), CD31 (e), CD119 (f), CD54 (g), and CD45 (h).

Supplementary Figure 3. Phenotypic Characteristics of ECs Cultured in the Presence of MVs Produced by NK Cells of the NK-92 Cell Line. 1) Distribution of ECs in a FSC/SSC histogram; 2) Isotypic control; 3) Distribution of ECs treated with antibodies to VEGFR1 (a), VEGFR2 (b), CD34 (c), CD105 (d), CD31 (e), CD119 (f), CD54 (g), and CD45 (h).

Supplementary Figure 4: Electrophoregrams of Lysates of Cells of the NK-92 Cell Line (Cells) and their MVs (MVs).

Supplementary Figure 5. ECs Incubated with MVs Produced by NK Cells of the NK-92 Cell Line. 1) Distribution of ECs in a FSC/SSC histogram; 2) Negative control, (a) distribution of unstimulated ECs in a FITC/FSC histogram, (b) FITC histogram; 3) (a) ECs treated with MVs derived from unstimulated NK cells in a FITC/FSC histogram, (b) FITC histogram; 4) (a) ECs treated with MVs derived from NK cells pretreated with CFSE 5 μm in a FITC/FSC histogram, (b) FITC histogram; 5) (a) ECs treated with MVs derived from NK cells pretreated with CFSE 50 μm in a FITC/FSC histogram, (b) FITC histogram.

Supplementary Figure 6. Representative immunoblots showing the content of ERK1/2, AKT in cells of the NK-92 cell line and their MVs. (1 – MVs derived from cells of the NK-92 cell line, 2 – cells of the NK-92 cell line).