РЕГУЛЯЦИЯ ЦИТОКИНОВОГО ПРОФИЛЯ NK-КЛЕТОК ФАКТОРАМИ МИКРООКРУЖЕНИЯ, ХАРАКТЕРНЫМИ ДЛЯ БЕРЕМЕННОСТИ

Гребенкина П.В.¹, Михайлова В.А.¹, Беспалова О.Н.¹, Сельков С.А.^{1, 2}, Соколов Д.И.^{1, 2}

¹ ФГБНУ «Институт акушерства, гинекологии и репродуктологии имени Д.О. Отта», Санкт-Петербург, Россия ² ФГБОУ ВО «Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова» Министерства здравоохранения РФ, Санкт-Петербург, Россия

Резюме. Децидуальные NK-клетки обладают отличными фенотипическими и функциональными характеристиками по сравнению с периферическими NK-клетками. Однако механизмы, лежащие в основе развития этих уникальных свойств, остаются малоизученными. Предполагается, что клетки макроокружения оказывают как прямое, так и косвенное влияние на NK-клетки в матке, модулируя их уровень «агрессии» по отношению к тканям плода, включая клетки трофобласт.

Одним из механизмов регуляции NK-клеток является выделение цитокинов. Трофобласты, как и другие элементы микроокружения, продуцируют цитокины. Связываясь с рецепторами на поверхности клеток-мишеней, они изменяют поведение NK-клеток. В результате NK-клетки сами могут выделять цитокины, которые, в свою очередь, влияют на поведение других клеток. Как упоминалось ранее, недостаточно данных о причинах и механизмах, лежащих в основе изменений характеристик NK-клеток в матке. Тем не менее эти данные могут стать основой для создания более точной клеточной модели взаимодействия между клетками плода и иммунной системой матери. Кроме того, они могут послужить основой для разработки инструментов диагностики репродуктивных проблем.

Целью исследования – изучение цитокинового профиля NK-клеток (продукция TNFα, TGF-β, IFNγ, RANTES, IL-10, VEGF) под влиянием цитокинов, связанных с беременностью, – TNFα, IFNγ, TGF-β1, IL-15, IL-18 или IL-10.

Уровни этих цитокинов в кондиционированных средах, полученных после культивирования NK-клеток, были измерены с помощью проточной цитометрии. Было обнаружено, что TGF-β1, секретируемый клетками трофобласта, обладает способностью регулировать цитокиновый профиль NK-клеток. Под его воздействием уровни IFNγ, IL-10 и RANTES в средах, полученных из культуры NK-клеток, были снижены.

Основываясь на этих результатах, можно сделать вывод, что существует система, которая контролирует активность NK-клеток через сеть цитокинов. Эти данные указывают на потенциальную возможность использования TGF-β1 для моделирования взаимодействия между NK-клетками и трофобластами *in vitro*.

Адрес для переписки:	Address for correspondence:
Гребенкина Полина Владимировна	Polina V. Grebenkina
$\hat{\Phi}$ ГБНУ «Институт акушерства, гинекологии	D. Ott Research Institute of Obstetrics,
и репродуктологии имени Д.О. Отта»	Gynecology and Reproductology
199034, Россия, Санкт-Петербург,	3 Mendeleevskaya Line
Менделеевская линия, д. 3.	St. Petersburg
Тел.: 8 (905) 802-85-58.	199034 Russian Federation
E-mail: grebenkinap@gmail.com	<i>Phone:</i> +7 (905) 802-85-58.
	E-mail: grebenkinap@gmail.com

Ключевые слова: NK-клетки, цитокины, беременность, трофобласт, TGF-β, межклеточные взаимодействия, микроокружение

Образец цитирования:

П.В. Гребенкина, В.А. Михайлова, О.Н. Беспалова, С.А. Сельков, Д.И. Соколов «Регуляция цитокинового профиля NK-клеток факторами микроокружения, характерными для беременности» // Медицинская иммунология, 2025. Т. 27, № 2. С. 445-450. doi: 10.15789/1563-0625-ROT-3061

© Гребенкина П.В. и соавт., 2025 Эта статья распространяется по лицензии Creative Commons Attribution 4.0

For citation:

P.V. Grebenkina, V.A. Mikhailova, O.N. Bespalova, S.A. Selkov, D.I. Sokolov "Regulation of the cytokine profile of NK cells by the microenvironment factors typical for pregnancy", Medical Immunology (Russia)/Meditsinskaya Immunologiya, 2025, Vol. 27, no. 2, pp. 445-450. doi: 10.15789/1563-0625-ROT-3061

© Grebenkina P.V. et al., 2025 The article can be used under the Creative Commons Attribution 4.0 License **DOI:** 10.15789/1563-0625-ROT-3061

REGULATION OF THE CYTOKINE PROFILE OF NK CELLS BY THE MICROENVIRONMENT FACTORS TYPICAL FOR PREGNANCY

Grebenkina P.V.^a, Mikhailova V.A.^a, Bespalova O.N.^a, Selkov S.A.^{a, b}, Sokolov D.I.^{a, b}

^a D. Ott Research Institute of Obstetrics, Gynecology and Reproductology, St. Petersburg, Russian Federation ^b First St. Petersburg State I. Pavlov Medical University, St. Petersburg, Russian Federation

Abstract. Decidual NK cells exhibit distinct phenotypic and functional characteristics as compared to peripheral NK cells. However, the mechanisms underlying development of these unique properties remain poorly understood. The cells in microenvironment are known to exert both direct and indirect influence on NK cells within uterus, modulating their level of "aggressiveness" towards fetal tissues, including trophoblasts. Cytokine release presents a remote regulatory tool for the NK cells. Trophoblasts produce cytokines like as other components of the microenvironment. These cytokines bind the receptors on surface of target cells thud changing the behavior of NK cells. As a result, NK cells may release the own cytokines, which, in turn, influence the behavior of other cells. As mentioned above, there is a lack of data on causes and mechanisms behind the changes in characteristics of NK cells in uterus. Nevertheless, this data can lay the foundation for designing a more accurate cellular model of interactions between fetal cells and maternal immune system. Moreover, it may serve as a basis for developing diagnostic tools for reproductive issues.

The aim of our study was to investigate changes in cytokine profile of NK cells, in particular, their production of TNF α , TGF- β , IFN γ , RANTES, IL-10, and VEGF under the influence of cytokines associated with pregnancy, i.e., TNF α , IFN γ , TGF- β 1, IL-15, IL-18, or IL-10. The levels of these cytokines in the culture media conditioned by NK cells were measured using flow cytometry. TGF- β 1, produced by trophoblasts was found to have the ability of regulating cytokine secretion by NK cells. The levels of IFN γ , IL-10, and RANTES in the media derived from NK cell culture have been decreased under its influence.

On the basis of these findings, one may propose the existence of a regulatory system that controls activity of NK cells via the cytokine network. These data suggest a potential for using TGF- β 1 to model *in vitro* interactions between NK cells and trophoblasts.

Keywords: NK cells, cytokines, pregnancy, trophoblast, TGF-\beta, intercellular communication, microenvironment

This study was funded by the project – "Development of diagnostic criteria for predicting and overcoming reproductive losses", head O.N. Bespalova. State registration number 122041500061-8.

Introduction

NK cells are found in peripheral blood, but there are also local populations of these cells with special properties. One such population is decidual NK cells, which are found in the uteroplacental contact zone. These cells have a different phenotype and function compared to NK cells in peripheral blood [6].

One of the mechanisms of long-distance regulation of natural killer (NK) cells is cytokine secretion. The primary population of cells that influence NK cells is trophoblasts, but other cells such as uterine macrophages and endometrial cells also produce cytokines in the context of maternalfetal interaction. Cytokines bind to receptors on cell membranes, triggering signaling cascades that result in changes in NK cell function, including cytokine secretion, which in turn allows NK cells to influence other cells. The cytokines under consideration in this study were detected in the region of uterine-placental interaction – IL-15, IL-18, IL-10, TGF- β , TNF α , IFN γ , RANTES VEGF [4, 7, 8, 15]. Data on possible uterine-placental cells that produce these cytokines are presented in Table 1.

The intricate molecular mechanisms governing the regulation of natural killer (NK) cell functions by pregnancy-specific cytokines remain underexplored. Therefore, the objective of this study was to elucidate the nature of alterations in the cytokine profile of NK cells in response to microenvironmental cues.

Materials and methods

Cell cultures

This study was performed using the NK-92 cell line (ATCC, USA), which reflect the main characteristics of and natural killer cells, respectively. The cells

Cytokine	Cell-producer	Effect
IL-15	Trophoblast cells, endometrial cells, placental macrophages	Regulation of effector functions of NK cells, stimulation of invasion and proliferation of trophoblast cells
IL-18	Decidual stromal cells	Induces cytotoxicity proteins expression by uterine, natural killer (uNK) cells
IL-10	NK cells, trophoblast cells	Inducing the effector functions of NK cells, stimulating the expression of the protective molecule HLA-G on the surface of trophoblast cells, anti-invasive effect on trophoblast cells
TGF-β	NK cells, trophoblast cells, T regulatory cells	Reduction of cytotoxicity of NK cells, regulation of trophoblast cell invasion
TNFα	NK cells	Enhancement of effector functions of NK cells, invasive effect on trophoblast cells
IFNγ	NK cells, trophoblast cells	Enhancement of effector functions of NK cells, inhibition of trophoblast cell invasion
RANTES	NK cells, trophoblast cells	Stimulation of NK cell proliferation and migration, increased migration and invasion of trophoblast cells
VEGF	Decidual NK cells, endometrial cells	Regulation of vascular growth and formation at the uteroplacental contact boundary. Promotes the acquisition of an endothelial phenotype by the extracellular trophoblast, which ensures the successful embedding of the trophoblast into the experimental vessel model

TABLE 1. LIST OF CYTOKINES ANALYZED IN THE RESEARCH AND THEIR EFFECTS

were cultured together with the manufacturer's recommendations in a humid environment at 37 °C, 5% CO₂. The viability of the cells was controlled by trypan blue exclusion, it was $95\pm3.4\%$.

Cytokines

The following inductors were used: $TNF\alpha$ (50 U/mL), $IFN\gamma$ (1000 U/mL), $TGF-\beta1$ (5 ng/mL), IL-15 (10 ng/mL), IL-18 (10 ng/mL), IL-10 (10 ng/mL) (R&D, USA). These concentrations were chosen according to the concentrations in human biological fluids, including in the area of uteroplacental contact.

NK-92 cells were added to a part of the wells in 100 μ L of a medium of 20,000 cells. IL-2 (500 U/mL) was added to all wells (LLC "Biotech", Russia), cytokines were added to some of the wells and then cells were cultured for 96 hours in the presence of TNF α , IFN γ , TGF- β 1, IL-15, IL-18, or IL-10. After that, the analysis of their cytokine production was conducted. Four experiments were conducted with two technical repetitions in each.

Analysis of cytokine level in conditioned media obtained after cultivation of NK-92 cells

After cultivation for 96 hours in the presence of cytokines, NK-92 cells were centrifuged for 5 minutes at 2500 g, the supernatants (hereinafter "conditioned media", CM) were frozen at -20 °C and stored until the study. Then, the level of IL-10, RANTES, VEGF, TNF α , IFN γ and TGF- β 1 in CM was evaluated using commercial kits for CBA (Cytometric Bead Array) (BD, USA) and a flow cytofluorimeter

BD FACSCanto II (BD, USA) according to the manufacturer's instructions.

Results and discussion

The content of TNF α , TGF- β , IFN γ , RANTES, IL-10, and VEGF was evaluated in CM obtained after cultivation of NK-92 cells for 96 hours in the presence of cytokines. We found that NK-92 cells on the baseline level secreted IFN γ , IL-10 and RANTES and in low concentrations TGF- β , but did not secrete VEGF and TNF α (Figure 1).

It has previously been shown that NK-92 cells are capable of secreting VEGF [13], TNF α [5]. During the 96-hour incubation period, it is likely that secreted cytokines are internalized by NK cells in order to regulate their properties. However, in order to confirm this hypothesis, further analysis will be required to determine the intracellular content of cytokines. The content of IFN γ , RANTES, and IL-10 in CM was reduced after cultivation of NK-92 cells in the presence of TGF- β 1 when compared with the content of these cytokines after cultivation without inducers (Figure 1).

TGF- β 1, a major cytokine secreted by trophoblast cells, exerts an immunosuppressive influence [11], nonetheless, it has been demonstrated that in certain instances, it may enhance the cytotoxic potency of cytotoxic cells [9]. The pro-inflammatory cytokine IFN γ enhances the effector functions of NK cells [12], has an inhibitory effect on trophoblast cells, reducing their ability to invade [1]. IL-10 it can



Figure 1. Content of IFN γ (A), IL-10 (B), RANTES (C), VEGF (D), TNF α (E), and TGF- β (F) in CM after cultivation of NK-92 cells in the presence of various cytokines (indicated on the abscissa axis) for 96 hours: NI – NK-92 cells without inductors Note. Significance of differences: *, p < 0.05; **, p < 0.01; ***, p < 0.001.

also serve as an enhancer of the effector capabilities of NK cells [3], elicit the secretion of IFN γ , thereby augmenting the cytotoxic capacity [2]. Moreover, the cytokine exerts an anti-invasive and pro-apoptotic influence on trophoblast cells [10]. Accordingly, inhibition of IFN-y and IL-10 secretion by the action of TGF- β 1 may be a mechanism used by trophoblast cells to establish a tolerant microenvironment for a semiallegenic fetus.

Nonetheless, this notion does not align with the reduction in RANTES secretion upon TGF- β 1 treatment. On the contrary, it has been observed that RANTES serves as a stimulant for the proliferation and migration of NK cells [14], this is a piece of evidence that supports the hypothesis of fostering tolerance towards fetal cells. Nonetheless, it should be noted that cytokine also serves as a stimulant for the migration and infiltration of trophoblast cells [4].

Consequently, a reduction in the production of NK cells hinders the development of fetal cells. This observation may suggest the existence of a regulatory mechanism employed by trophoblast cells to prevent their own over-invasion.

Conclusion

In the presence of TGF- β 1, NK cells decrease the production of IFN γ , IL-10 and RANTES. This finding may indicate that trophoblast cells and other cells in the microenvironment, producing TGF- β 1, can alter NK cell function, creating an optimal environment for fetal development. The data obtained reflect processes occurring between cells of the maternal and fetal immune systems during pregnancy and could serve as a basis for developing tests to diagnose causes of reproductive pathologies resulting from disruptions in intercellular communication.

References

1. Bačenková D., Trebuňová M., Čížková D., Hudák R., Dosedla E., Findrik-Balogová A., Živčák J. *In vitro* model of human trophoblast in early placentation. *Biomedicines*, 2022, Vol. 10, no. 4, 904. doi: 10.3390/biomedicines10040904.

2. Burgess S.J., Marusina A.I., Pathmanathan I., Borrego F., Coligan J.E. IL-21 down-regulates NKG2D/DAP10 expression on human NK and CD8⁺ T cells. *J. Immunol.*, 2006, Vol. 176, no. 3, pp. 1490-1497.

3. Cai G., Kastelein R.A., Hunter C.A. IL-10 enhances NK cell proliferation, cytotoxicity and production of IFN-gamma when combined with IL-18. *Eur. J. Immunol.*, 1999, *Vol. 29, no. 9, pp. 2658-2665.*

4. Fujiwara H., Higuchi T., Sato Y., Nishioka Y., Zeng B.X., Yoshioka S., Tatsumi K., Ueda M., Maeda M. Regulation of human extravillous trophoblast function by membrane-bound peptidases. *Biochim. Biophys. Acta*, 2005, Vol. 1751, no. 1, pp. 26-32.

5. Gómez-Lomelí P., Bravo-Cuellar A., Hernández-Flores G., Jave-Suárez L.F., Aguilar-Lemarroy A., Lerma-Díaz J.M., Domínguez-Rodríguez J.R., Sánchez-Reyes K., Ortiz-Lazareno P.C. Increase of IFN-γ and TNF-γ production in CD107a + NK-92 cells co-cultured with cervical cancer cell lines pre-treated with the HO-1 inhibitor. *Cancer Cell Int., 2014, Vol. 14, no. 1, 100.* doi: 10.1186/s12935-014-0100-1.

6. Hanna J., Goldman-Wohl D., Hamani Y., Avraham I., Greenfield C., Natanson-Yaron S., Prus D., Cohen-Daniel L., Arnon T.I., Manaster I., Gazit R., Yutkin V., Benharroch D., Porgador A., Keshet E., Yagel S., Mandelboim O. Decidual NK cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med*, 2006, *Vol. 12, no. 9, pp. 1065-1074.*

7. Klimkiewicz-Blok D., Florjanski J., Zalewski J., Blok R. Analysis of the concentrations of interleukin 15 in amniotic fluid in the second and the third trimesters of pregnancy. *Adv. Clin. Exp. Med.*, 2012, Vol. 21, no. 1, pp. 75-79.

8. Klimkiewicz-Blok D., Florjanski J., Zalewski J., Blok R. Analysis of the concentrations of interleukin 18 in amniotic fluid in the second and the third trimesters of pregnancy. *Adv. Clin. Exp. Med.*, 2013, Vol. 22, no. 5, pp. 699-703.

9. Peters C., Meyer A., Kouakanou L., Feder J., Schricker T., Lettau M., Janssen O., Wesch D., Kabelitz D. TGF-beta enhances the cytotoxic activity of Vdelta2 T cells. *Oncoimmunology, 2019, Vol. 8, no. 1, e1522471.* doi: 10.1080/2162402X.2018.1522471.

10. Roth I., Fisher S.J. IL-10 is an autocrine inhibitor of human placental cytotrophoblast MMP-9 production and invasion. *Dev. Biol.*, 1999, Vol. 205, no. 1, pp. 194-204.

11. Viel S., Marcais A., Guimaraes F.S., Loftus R., Rabilloud J., Grau M., Degouve S., Djebali S., Sanlaville A., Charrier E., Bienvenu J., Marie J.C., Caux C., Marvel J., Town L., Huntington N.D., Bartholin L., Finlay D., Smyth M.J., Walzer T. TGF-beta inhibits the activation and functions of NK cells by repressing the mTOR pathway. *Sci. Signal.*, 2016, *Vol. 9, no. 415, ra19.* doi: 10.1126/scisignal.aad1884.

12. Wang R., Jaw J.J., Stutzman N.C., Zou Z., Sun P.D. Natural killer cell-produced IFN-gamma and TNF-alpha induce target cell cytolysis through up-regulation of ICAM-1. *J. Leukoc. Biol.*, 2012, Vol. 91, no. 2, pp. 299-309.

13. Wang X., Liang C., Xia W., Guo C., Niu Z., Zhu W., Zhang H. VEGF165b augments NK92 cytolytic activity against human K562 leukemia cells by upregulating the levels of perforin and granzyme B via the VEGR1-PLC pathway. *Mol. Immunol.*, 2020, Vol. 128, pp. 41-46.

14. Weiss I.D., Shoham H., Wald O., Wald H., Beider K., Abraham M., Barashi N., Galun E., Nagler A., Peled A. Ccr5 deficiency regulates the proliferation and trafficking of natural killer cells under physiological conditions. *Cytokine*, 2011, Vol. 54, no. 3, pp. 249-257.

15. Zhang C., Tian Z. NK cell subsets in autoimmune diseases. J. Autoimmun., 2017, Vol. 83, pp. 22-30.

Авторы:

Гребенкина П.В. — младший научный сотрудник ФГБНУ «Институт акушерства, гинекологии и репродуктологии имени Д.О. Отта», Санкт-Петербург, Россия

Михайлова В.А. — д.б.н., старший научный сотрудник лаборатории межклеточных взаимодействий ФГБНУ «Институт акушерства, гинекологии и репродуктологии имени Д.О. Отта», Санкт-Петербург, Россия

Authors:

Grebenkina P.V., Junior Research Associate, D. Ott Research Institute of Obstetrics, Gynecology and Reproductology, St. Petersburg, Russian Federation

Mikhailova V.A., PhD, MD (Biology), Senior Research Associate, Laboratory of Intercellular Interactions, D. Ott Research Institute of Obstetrics, Gynecology and Reproductology, St. Petersburg, Russian Federation Беспалова О.Н. – д.м.н., заместитель директора по научной работе ФГБНУ «Институт акушерства, гинекологии и репродуктологии имени Д.О. Отта», Санкт-Петербург, Россия

Сельков С.А. — д.м.н., профессор, заслуженный деятель науки РФ, заведующий отделом иммунологии и межклеточных взаимодействий ФГБНУ «Научноисследовательский институт акушерства, гинекологии и репродуктологии имени Д.О. Отта»; профессор кафедры иммунологии ФГБОУ ВО «Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова» Министерства здравоохранения РФ, Санкт-Петербург, Россия

Соколов Д.И. — д.б.н., доцент, заведующий лабораторией межклеточных взаимодействий ФГБНУ «Научно-исследовательский институт акушерства, гинекологии и репродуктологии имени Д.О. Отта»; профессор кафедры иммунологии ФГБОУ ВО «Первый Санкт-Петербургский государственный медицинский университет имени академика И.П. Павлова» Министерства здравоохранения РФ, Санкт-Петербург, Россия

Поступила 29.07.2024 Отправлена на доработку 30.07.2024 Принята к печати 14.09.2024 **Bespalova O.N.,** PhD, MD (Medicine), Deputy Director for Research, D. Ott Research Institute of Obstetrics, Gynecology and Reproductology, St. Petersburg, Russian Federation

Selkov S.A., PhD, MD (Medicine), Professor, Honored Scientist of the Russian Federation, Head, Department of Immunology and Intercellular Communications, D. Ott Research Institute of Obstetrics, Gynecology and Reproductology; Professor, Department of Immunology, First St. Petersburg State I. Pavlov Medical University, St. Petersburg, Russian Federation

Sokolov D.I., PhD, MD (Biology), Associate Professor, Head, Laboratory of Intercellular Communications, D. Ott Research Institute of Obstetrics, Gynecology and Reproductology; Professor, Department of Immunology, First St. Petersburg State I. Pavlov Medical University, St. Petersburg, Russian Federation

Received 29.07.2024 Revision received 30.07.2024 Accepted 14.09.2024