

## **ФУНКЦИОНАЛЬНОЕ ИСТОЩЕНИЕ CD4<sup>+</sup>T-КЛЕТОК У ВИЧ/ВГС-КОИНФИЦИРОВАННЫХ ПАЦИЕНТОВ, ПОЛУЧАЮЩИХ ВЫСОКОАКТИВНУЮ АНТИРЕТРОВИРУСНУЮ ТЕРАПИЮ**

**Власова В.В., Королевская Л.Б., Логинова О.А., Шмагель Н.Г.,  
Сайдакова Е.В.**

*Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал  
ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук»,  
г. Пермь, Россия*

**Резюме.** ВГС-коинфекция широко распространена среди ВИЧ-инфицированных пациентов. В России до 50% ВИЧ-позитивных больных коинфицированы ВГС. Применение высокоактивной антиретровирусной терапии (ВААРТ), в большинстве случаев, приводит к подавлению репликации ВИЧ и восстановлению иммунной системы ВИЧ-инфицированных пациентов. Однако ВГС-коинфекция препятствует эффективному восстановлению CD4<sup>+</sup>T-клеток и повышает риск заболеваемости и смерти ВИЧ-инфицированных больных, получающих ВААРТ. Известно, что скорость прогрессии ВИЧ-инфекции и эффективность восстановления иммунной системы на фоне приема ВААРТ в значительной мере зависят от уровня активации иммунной системы и степени истощения CD4<sup>+</sup>T-клеток. Целью настоящей работы было определение уровня активации и истощения, а также цитокин-продуцирующей функции CD4<sup>+</sup>T-клеток, полученных из крови получающих ВААРТ ВИЧ/ВГС-коинфицированных и ВИЧ-моноинфицированных больных. В исследование были включены ВИЧ/ВГС-коинфицированные (n = 11) и ВИЧ-моноинфицированные (n = 10) пациенты, получающие ВААРТ более двух лет. В контрольную группу вошли 10 добровольцев без признаков ВИЧ- и ВГС-инфекций. Было установлено, что у ВИЧ/ВГС-коинфицированных пациентов, в сравнении со здоровыми людьми, повышены следующие показатели: доля активированных CD38<sup>+</sup>HLA-DR<sup>+</sup> CD4<sup>+</sup>T-лимфоцитов (p < 0,05), уровень истощения CD4<sup>+</sup>T-клеток, определенный по плотности экспрессии TIMT на поверхности каждой клетки (p < 0,05), и число CD4<sup>+</sup>T-лимфоцитов, способных производить интерферон-гамма (IFN $\gamma$ ) после активации (p < 0,05). Относительное число IFN $\gamma$ -продуцирующих CD4<sup>+</sup>T-лимфоцитов в крови доноров позитивно коррелировало с долей активированных CD4<sup>+</sup>T-клеток (R = 0,514, p < 0,01). Важно отметить, что, несмотря на большое ко-

### **Адрес для переписки:**

Власова Виолетта В.  
Институт экологии и генетики микроорганизмов  
Уральского отделения Российской академии наук  
614081, Россия, г. Пермь, ул. Голева, 13  
Тел.: 8 (996) 324-12-01.  
Факс: 8 (342) 280-83-34.  
E-mail: violetbaudelaire73@gmail.com

### **Address for correspondence:**

Violetta V. Vlasova  
Institute of Ecology and Genetics of Microorganisms  
13 Golev St  
Perm  
614081 Russian Federation  
Phone: +7 (996) 324-12-01.  
Fax: +7 (342) 280-83-34.  
E-mail: violetbaudelaire73@gmail.com

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личество  $\text{IFN}\gamma$ -продуцирующих  $\text{CD4}^+\text{T}$ -лимфоцитов, средняя продукция этого цитокина в  $\text{CD4}^+\text{T}$ -клетках ВИЧ/ВГС-коинфицированных больных была существенно ниже, чем у здоровых субъектов ( $p < 0,05$ ). Продукция  $\text{IFN}\gamma$  в  $\text{CD4}^+\text{T}$ -лимфоцитах не зависела от степени их активации ( $p > 0,05$ ). Негативная корреляционная связь была установлена между содержанием  $\text{IFN}\gamma$  и уровнем истощения  $\text{CD4}^+\text{T}$ -клеток ( $R = -0,400$ ,  $p < 0,05$ ). Показатели истощения  $\text{CD4}^+\text{T}$ -лимфоцитов также обратно коррелировали с содержанием  $\text{CD4}^+\text{T}$ -клеток в крови ( $R = -0,598$ ,  $p < 0,01$ ). Полученные сведения позволяют предположить, что ВГС-коинфекция приводит к выраженному функциональному истощению  $\text{CD4}^+\text{T}$ -клеток и тем самым может отягощать течение ВИЧ-инфекции у пациентов, принимающих ВААРТ.

**Ключевые слова:** ВИЧ-инфекция, гепатит С,  $\text{CD4}^+\text{T}$ -лимфоциты, иммунная активация, иммунное истощение, продукция цитокинов

## FUNCTIONAL EXHAUSTION OF $\text{CD4}^+\text{T}$ CELLS IN HIV/HCV COINFECTED HAART-TREATED PATIENTS

Vlasova V.V., Korolevskaya L.B., Loginova O.A., Shmagel N.G., Saidakova E.V.

*Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation*

**Abstract.** Infection with hepatitis C virus (HCV) is common among HIV-positive patients, with up to 50% of them being coinfecting in Russia. While highly active antiretroviral therapy (HAART) suppresses HIV replication and restores the immune system of HIV-infected subjects, HCV coinfection interferes with  $\text{CD4}^+\text{T}$  cell regeneration and increases the risk of patients' morbidity and mortality. During HAART, HIV-infection progression and the immune system restoration efficiency largely depend on immune activation and  $\text{CD4}^+\text{T}$  cell exhaustion. This study determined the level of activation, exhaustion, and cytokine production in  $\text{CD4}^+\text{T}$  cells obtained from the peripheral blood of HAART-treated HIV/HCV coinfecting and HIV mono-infected subjects. The study comprised 11 HIV/HCV coinfecting individuals and 10 HIV mono-infected patients receiving HAART for more than two years, with a control group of 10 volunteers without the signs of HIV or HCV infections. Compared with healthy controls, HIV/HCV coinfecting patients had an increased frequency of activated  $\text{CD38}^+\text{HLA-DR}^+$   $\text{CD4}^+\text{T}$  lymphocytes ( $p < 0.05$ ), a higher level of  $\text{CD4}^+\text{T}$  cell exhaustion determined according to the TIGIT expression density per cell ( $p < 0.05$ ), and a greater proportion of interferon-gamma ( $\text{IFN}\gamma$ )-producing  $\text{CD4}^+\text{T}$  lymphocytes following activation ( $p < 0.05$ ). The frequency of  $\text{IFN}\gamma$ -producing  $\text{CD4}^+\text{T}$  cells in the donors' blood positively correlated with the proportion of activated  $\text{CD4}^+\text{T}$  cells ( $R = 0.514$ ,  $p < 0.01$ ). Despite having a large number of  $\text{IFN}\gamma$ -producing  $\text{CD4}^+\text{T}$  lymphocytes, the HIV/HCV coinfecting patients' average production of  $\text{IFN}\gamma$  by  $\text{CD4}^+\text{T}$  cells was significantly lower than that in healthy controls ( $p < 0.05$ ). The  $\text{IFN}\gamma$  production in  $\text{CD4}^+\text{T}$  lymphocytes did not depend on activation ( $p > 0.05$ ). However, a negative correlation was established between the  $\text{IFN}\gamma$  production and the level of  $\text{CD4}^+\text{T}$  cell exhaustion ( $R = -0.400$ ,  $p < 0.05$ ). The latter was also found to inversely correlate with the  $\text{CD4}^+\text{T}$  cell counts in the donors' peripheral blood ( $R = -0.598$ ,  $p < 0.01$ ). These data suggest that HCV coinfection leads to pronounced functional exhaustion of  $\text{CD4}^+\text{T}$  cells and may aggravate the course of HIV-infection in patients receiving HAART.

**Keywords:** HIV-infection, hepatitis C,  $\text{CD4}^+\text{T}$  cells, immune activation, immune exhaustion, cytokine production

This work was carried out within the framework of the State assignment #121112500044-9.

### Introduction

Due to shared transmission routes, hepatitis C virus (HCV) coinfection has become prevalent among

HIV-positive individuals. Worldwide, approximately 25% of HIV-infected patients are coinfecting with HCV [7], while in Russia the proportion of HIV/HCV coinfecting subjects goes up to 50% [15]. The majority of HIV-positive people receive highly active antiretroviral therapy (HAART), which suppresses

HIV replication and leads to the immune system regeneration defined by the increase in the peripheral blood CD4<sup>+</sup>T cell counts [6]. However, in HIV/HCV coinfecting people receiving HAART, hepatitis C infection interferes with CD4<sup>+</sup>T cell recovery [14]. Furthermore, compared with HAART-treated HIV-monoinfected subjects, HIV/HCV coinfecting patients have a higher risk of developing non-AIDS-associated diseases [10], opportunistic infections [3], and death [1]. The mechanisms behind the negative effect of hepatitis C on the course of treated HIV-infection remain poorly understood.

Immune activation and CD4<sup>+</sup>T cell exhaustion are critical for the natural history of HIV-infection, affecting both the pace of the disease progression and the immune restoration efficiency during HAART [9, 11]. In treated HIV-infected individuals, HCV coinfection has been shown to significantly increase the chronic immune activation and systemic inflammation levels [12]. However, it is unclear if it affects the CD4<sup>+</sup>T cells' functional state. Therefore, the purpose of this study was to determine the level of CD4<sup>+</sup>T cells' activation and exhaustion, as well as the cytokine-producing function of these lymphocytes obtained from the blood of HAART-treated HIV/HCV coinfecting and HIV monoinfected patients.

## Materials and methods

The work plan was approved by the ethics committee of the Perm Regional Center for the Prevention and Control of AIDS and Infectious Diseases (committee registration number IRB00008964). The study included HIV-infected patients receiving HAART for more than two years (viral load < 50 copies/mL): 1) HIV/HCV coinfecting subjects (n = 11); and 2) HIV-monoinfected patients (n = 10). The healthy control group (HC) comprised 10 volunteers without HIV or HCV infections.

Blood was collected from the cubital vein into Vacutainer tubes containing ethylenediaminetetraacetic acid. CD4<sup>+</sup>T lymphocytes count was assessed using the commercial BD Simultest™ IMK-Lymphocyte kit ("BD Biosciences", USA) with a CytoFLEX S flow cytometer ("Beckman Coulter", USA). The HIV viral load was determined using Versant HIV-1 RNA 3.0 assay kits by means of a Versant 440 analyzer ("Siemens", Germany).

Mononuclear cells were isolated by density centrifugation using DiaColl (1.077 g/mL, "Diaem", Russia). The samples were stored in liquid nitrogen in a medium containing 90% fetal bovine serum (FBS, "Biowest", France) and 10% dimethyl sulfoxide ("AppliChem", Germany). In a day of the study, the cells were thawed at +37 °C, washed

in 10 mL of complete culture medium (10% FCS, 100 U/mL penicillin, and 100 µg/mL streptomycin ("Sigma", USA) in RPMI-1640), and then in 10 mL of Dulbecco's phosphate buffered saline (DPBS; "Gibco", USA). The cytokine-producing function of CD4<sup>+</sup>T cells was assessed after 4 hours of incubation in complete culture medium containing phorbol 12-myristate-13-acetate (PMA), ionomycin, and brefeldin A ("BioLegend", USA).

Peripheral blood mononuclear cells were analyzed on CytoFLEX S flow cytometer ("Beckman Coulter", USA). Viable cells were identified by the absence of staining with the vital dye Zombie UV Fixable Viability Kit ("BioLegend", USA). Anti-CD3-BV605 and anti-CD4-PE antibodies ("BioLegend", USA) were used to identify CD4<sup>+</sup>T lymphocytes. Expression of the TIGIT exhaustion marker was assessed using anti-TIGIT-AF488 antibodies ("BioLegend", USA). Anti-IFNγ-APC antibodies ("BioLegend", USA) were utilized to study the production of interferon-γ (IFNγ). Activated cells were identified using anti-CD38-PE/Fire700 ("BioLegend", USA) and anti-HLA-DR-APC-R700 ("BD Biosciences", USA) antibodies.

Statistical analysis and graph plotting were performed using the "Statistica 6" software. The data are presented as medians, interquartile ranges (25–75 percentile), and 10–90% intervals. Significance of differences between groups was established based on the Mann–Whitney U test. Correlation analysis was performed by the r-Pearson linear correlation test.

## Results and discussion

All groups were similar according to age and gender (Table 1). The HIV-infection and HAART duration, HIV viral load, and CD4<sup>+</sup>T cell counts did not differ between the groups comprised of HIV-infected subjects. However, CD4<sup>+</sup>T cell counts in these groups were significantly reduced compared to HCs (p < 0.05).

In HIV/HCV coinfecting patients compared with healthy individuals, the proportion of activated (CD38<sup>+</sup>HLA-DR<sup>+</sup>) CD4<sup>+</sup>T lymphocytes was increased (Figure 1A; p < 0.05). Meanwhile, the difference between HIV monoinfected patients and healthy controls was not statistically significant (p > 0.05).

The level of exhaustion in CD4<sup>+</sup>T cells of HIV/HCV coinfecting subjects was also increased, as evidenced by the higher frequency of TIGIT<sup>+</sup>CD4<sup>+</sup>T lymphocytes (p < 0.05; data not shown) and the elevated expression of the inhibitory receptor TIGIT on the surface of these cells (Figure 1B; p < 0.05).

TABLE 1. CLINICAL CHARACTERISTICS OF HIV-INFECTED AND HEALTHY INDIVIDUALS

Characteristics	HIV/HCV coinfectd patients	HIV monoinfected patients	Healthy Controls (HC)
Number of enrollments	11	10	10
Age (years)	40.0* (37.5-42.0)	43.5 (37.5-44.8)	43.0 (37.0-45.8)
Females (%)	45	50	50
HIV-infection duration (years)	16.0 (10.0-19.0)	9.5 (5.0-16.0)	—
HAART duration (years)	8.0 (3.8-13.0)	5.0 (2.5-6.2)	—
HIV viral load (copies/mL)	< 50**	< 50	—
CD4 <sup>+</sup> T cell counts ( $\mu\text{L}^{-1}$ )	542 (405-663) pHC < 0.05	616 (480-763) pHC < 0.05	707 (646-880)

Note. \*, the data presented as medians and interquartile ranges. \*\*, the test-system sensitivity limit. HAART, highly active antiretroviral therapy. Significance of differences between groups was established based on the Mann–Whitney U test.

The number of CD4<sup>+</sup>T cells that produced IFN $\gamma$  upon activation with PMA and ionomycin was significantly higher in HIV/HCV-positive individuals as compared with healthy donors (Figure 2A;  $p < 0.05$ ). Across all study groups, the number of IFN $\gamma$ -producing CD4<sup>+</sup>T lymphocytes positively correlated with the relative number of activated (CD38<sup>+</sup>HLA-DR<sup>+</sup>) CD4<sup>+</sup>T cells (Figure 2B).

Importantly, although the number of IFN $\gamma$ -producing CD4<sup>+</sup>T lymphocytes in HIV/HCV coin-

fectd individuals was increased, the average production of IFN $\gamma$  in CD4<sup>+</sup>T cells of these patients was significantly lower than that of healthy subjects (Figure 3A;  $p < 0.05$ ). The IFN $\gamma$  production in CD4<sup>+</sup>T lymphocytes did not depend on the percent of activated cells. However, a negative correlation was established between IFN $\gamma$  production and the level of CD4<sup>+</sup>T cell exhaustion (Figure 3B).

Both phenotypic and functional indexes of CD4<sup>+</sup>T lymphocyte exhaustion correlated with

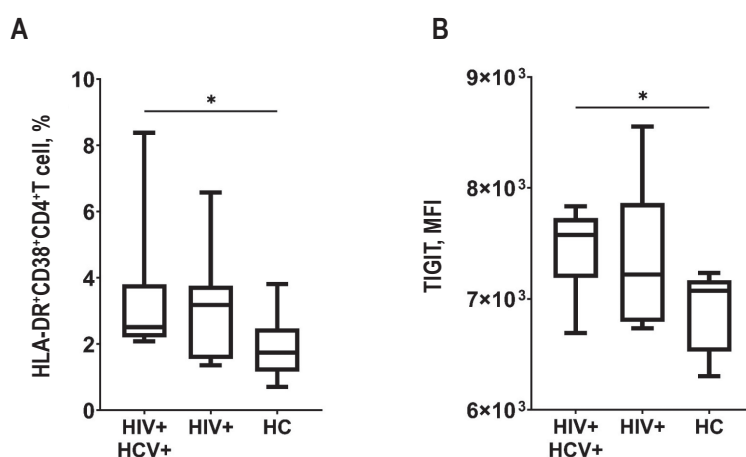
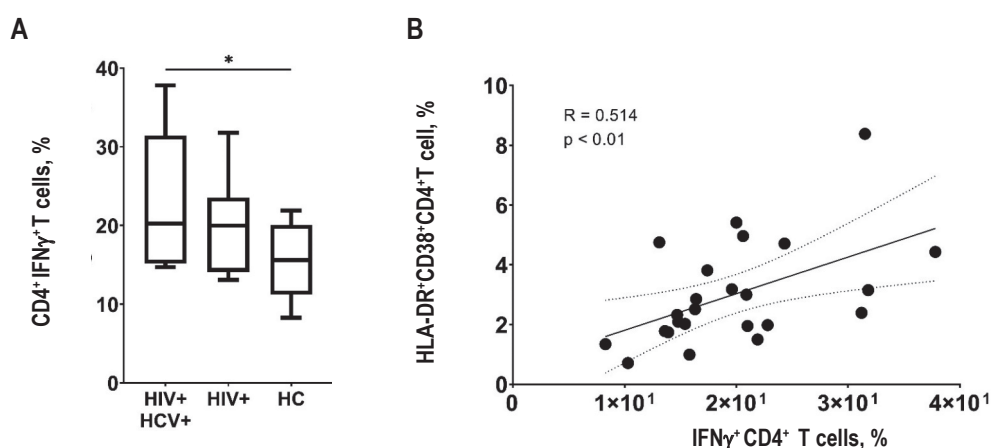


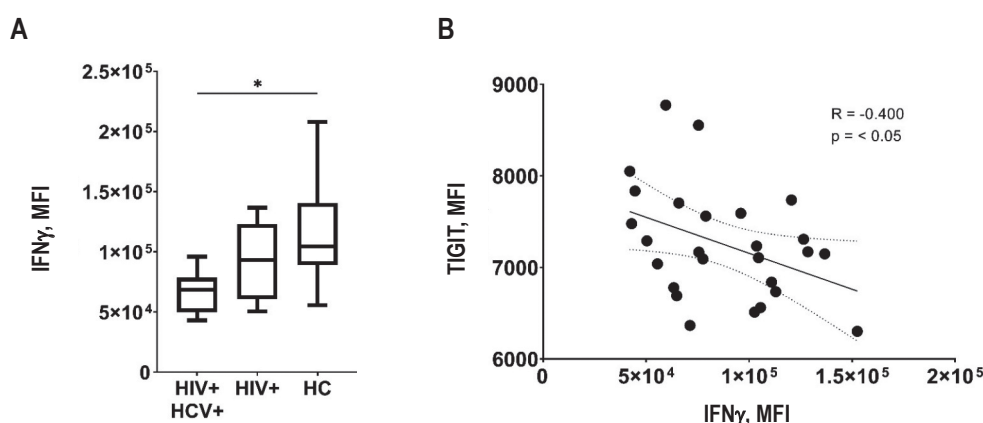
Figure 1. Level of CD4<sup>+</sup>T cell activation (A) and exhaustion (B) in HIV/HCV coinfectd people (HIV+HCV+), HIV monoinfected patients (HIV+) and healthy control (HC) subjects

Note. Medians (horizontal lines within rectangles), interquartile ranges (rectangles), and 10-90% intervals (vertical lines) are shown. \*,  $p < 0.05$  (Mann–Whitney U test). MFI, mean fluorescence intensity.



**Figure 2. Frequency of IFN $\gamma$  producing CD4<sup>+</sup>T lymphocytes (A) and its correlation with the relative number of activated CD4<sup>+</sup>T cells (B)**

Note. (A) Medians (horizontal lines within rectangles), interquartile ranges (rectangles), and 10-90% intervals (vertical lines) are shown. \*,  $p < 0.05$  (Mann–Whitney U test). (B) Individual donor values, regression line and 95% confidence intervals are shown (r-Pearson correlation test). "HIV+HCV+", HIV/HCV coinfecting patients; "HIV+", HIV mono-infected patients; "HC", healthy controls.



**Figure 3. Level of IFN $\gamma$  production (A) and its correlation with the degree of exhaustion (B) in CD4<sup>+</sup>T cells**

Note. As for Figure 2.

CD4<sup>+</sup>T cell counts in donors' blood. Specifically, CD4<sup>+</sup>T lymphocytes count was directly related to the production of IFN $\gamma$  ( $R = 0.455$ ,  $p < 0.05$ ) and inversely related to the proportion of exhausted CD4<sup>+</sup>T cells ( $R = -0.598$ ,  $p < 0.01$ ).

These data demonstrate that in HIV-infected patients, HCV coinfection is associated with a high level of CD4<sup>+</sup>T cells' activation and exhaustion. CD4<sup>+</sup>T lymphocytes in HIV/HCV coinfecting patients show both phenotypic (expression of the inhibitory receptor – TIGIT) and functional (decrease in IFN $\gamma$  production) signs of exhaustion. The frequency of exhausted CD4<sup>+</sup>T cells inversely correlate with the total number of CD4<sup>+</sup>T lymphocytes in donors' blood. These findings suggest that the negative impact of HCV on the course of HIV infection controlled

by HAART is due to the functional exhaustion of CD4<sup>+</sup>T cells.

Previously, researchers assessed the level of CD4<sup>+</sup>T cell exhaustion in HIV/HCV coinfecting patients by analyzing the relative number of lymphocytes expressing inhibitory receptor PD-1 [4]. The authors found that the frequency of PD1<sup>+</sup>CD4<sup>+</sup>T lymphocytes in HIV/HCV coinfecting patients was higher than that in HIV mono-infected subjects or healthy individuals. A noteworthy detail is that the surface expression of inhibitory receptors alone does not allow identifying truly exhausted CD4<sup>+</sup>T cells, as these molecules are also expressed by CD4<sup>+</sup>T lymphocytes that have received an activation signal [13]. However, exhausted and activated CD4<sup>+</sup>T cells differ significantly in terms of the



inhibitory receptors' expression level [5]. The amount of inhibitory receptors expressed on the surface of CD4<sup>+</sup>T cells in HIV/HCV coinfecting patients have not been reported in the literature yet. To account for this limitation, we assessed the level of CD4<sup>+</sup>T cells exhaustion by measuring the surface expression of the inhibitory receptor TIGIT, rather than by identifying TIGIT<sup>+</sup>CD4<sup>+</sup>T lymphocytes alone. Our findings show that the level of TIGIT expression on the surface of CD4<sup>+</sup>T lymphocytes is significantly higher in HIV/HCV coinfecting patients than in healthy individuals.

Determining the state of exhaustion in T cells requires assessing both their phenotype and function. TIGIT that was shown to be abundant on CD4<sup>+</sup>T cells of HIV/HCV coinfecting subjects inhibits the interaction between the costimulatory CD226 molecule and their common ligand, CD155 [8], leading to a decrease in T lymphocyte proliferation and cytokine production [5]. Previous studies have shown that in HIV-infected patients exhausted TIGIT<sup>+</sup>CD8<sup>+</sup>T cells are characterized by the reduced production of IFN $\gamma$ , TNF, and IL-2 [2]. However in HIV/HCV coinfecting patients, the cytokine-producing ability of the exhausted CD4<sup>+</sup>T cell pool has not been evaluated. In this study,

we have demonstrated for the first time that the IFN $\gamma$  production is reduced in CD4<sup>+</sup>T lymphocytes of HIV/HCV coinfecting patients receiving HAART. These results strongly suggest a high level of CD4<sup>+</sup>T cell exhaustion in patients coinfecting with HIV and HCV.

## Conclusion

An important finding of this study was the establishment of a link between the frequency of CD4<sup>+</sup>T lymphocytes expressing TIGIT, the level of IFN $\gamma$  production, and the amount of CD4<sup>+</sup>T cells in the patients' blood. CD4<sup>+</sup>T cell counts are a significant clinical indicator in HIV-infection as it determines the likelihood of developing non-AIDS-related diseases and death. Therefore, it can be assumed that the CD4<sup>+</sup>T cell exhaustion caused by HCV coinfection may adversely impact the course of HIV-infection in patients adherent to HAART.

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## References

1. Chen T.Y., Ding E.L., Seage III G.R., Kim A.Y. Meta-analysis: increased mortality associated with hepatitis C in HIV-infected persons is unrelated to HIV disease progression. *Clin Infect Dis*, 2009, Vol. 49, no. 10, pp. 1605-1615.
2. Chew G.M., Fujita T., Webb G.M., Burwitz B.J., Wu H.L., Reed J.S., Hammond K.B., Clayton, K.L., Ishii N., Abdel-Mohsen M., Liegler T., Mitchell B.I., Hecht F.M., Ostrowski M., Shikuma C.M., Hansen S.G., Maurer M., Korman A.J., Deeks S.G., Sacha J.B., Ndhlovu L.C. TIGIT marks exhausted T Cells, correlates with disease progression, and serves as a target for immune restoration in HIV and SIV infection. *PLoS Pathog.*, 2016, Vol. 12, no. 1, e1005349. doi: 10.1371/journal.ppat.1005349.
3. d'Arminio Monforte A., Cozzi-Lepri A., Castagna A., Antinori A., de Luca A., Mussini C., Caputo S.L., Arlotti M., Magnani G., Pellizzer G., Maggiolo F., Puoti M., Icona Foundation Study Group. Risk of developing specific AIDS-defining illnesses in patients coinfecting with HIV and hepatitis C virus with or without liver cirrhosis. *Clin. Infect. Dis.*, 2009, Vol. 49, no. 4, pp. 612-622.
4. Feuth T., Arends J.E., Fransen J.H., Nanlohy N.M., van Erpecum K.J., Siersema P.D., Hoepelman A.I.M., van Baarle D. Complementary role of HCV and HIV in T-cell activation and exhaustion in HIV/HCV coinfection. *PLoS One*, 2013, Vol. 8, no. 3, e59302. doi: 10.1371/journal.pone.0059302.
5. Freeman G.J., Wherry E.J., Ahmed R., Sharpe A.H. Reinvigorating exhausted HIV-specific T cells via PD-1-PD-1 ligand blockade. *J. Exp. Med.*, 2006, Vol. 203, no. 10, pp. 2223-2227.
6. Guihot A., Tubiana R., Breton G., Marcelin A.G., Samri A., Assoumou L., Goncalves E., Bricaire F., Costagliola D., Calvez V., Rouzioux C., Autran B., Katlama C., Carcelain G., ALT-ANRS CO-15 study group, DECAMUNE study group. Immune and virological benefits of 10 years of permanent viral control with antiretroviral therapy. *AIDS*, 2010, Vol. 24, no.4, pp. 614-617.
7. Hernandez M.D., Sherman K.E. HIV/hepatitis C coinfection natural history and disease progression. *Curr. Opin. HIV AIDS*, 2011, Vol. 6, no. 6, pp. 478-482.
8. Joller N., Kuchroo V.K. Tim-3, Lag-3, and TIGIT. *Curr. Top. Microbiol. Immunol.*, 2017, Vol. 410, pp. 127-156.
9. Okoye A.A., Picker L.J. CD4(+) T-cell depletion in HIV infection: mechanisms of immunological failure. *Immunol. Rev.*, 2013, Vol. 254, no. 1, pp. 54-64.
10. Operskalski E.A., Kovacs A. HIV/HCV co-infection: pathogenesis, clinical complications, treatment, and new therapeutic technologies. *Curr. HIV/AIDS Rep.*, 2011, Vol. 8, no. 1, pp. 12-22.

11. Rallón N., García M., García-Samaniego J., Cabello A., Álvarez B., Restrepo C., Nistal S., Górgolas M., Benito J.M. Expression of PD-1 and Tim-3 markers of T-cell exhaustion is associated with CD4 dynamics during the course of untreated and treated HIV infection. *PLoS One*, 2018, Vol. 13, no. 3, 0193829. DOI:10.1371/journal.pone.0193829
12. Shmagel K.V., Saidakova E.V., Shmagel N.G., Korolevskaya L.B., Chereshev V.A., Robinson J., Grivel J-C., Douek D.C., Margolis L., Anthony D.D., Lederman M.M. Systemic inflammation and liver damage in HIV/hepatitis C virus coinfection. *HIV Med.*, 2016, Vol. 17, no. 8, pp. 581-589.
13. Simon S., Labarriere N. PD-1 expression on tumor-specific T cells: Friend or foe for immunotherapy? *Oncoimmunology*, 2017, Vol. 7, no. 1, e1364828. doi: 10.1080/2162402X.2017.1364828.
14. Taye S., Lakew M. Impact of hepatitis C virus co-infection on HIV patients before and after highly active antiretroviral therapy: an immunological and clinical chemistry observation, Addis Ababa, Ethiopia. *BMC Immunol.*, 2013, Vol. 14, 23. doi: 10.1186/1471-2172-14-23.
15. Zatoloka P.A. Prevalence of concomitant pathology in HIVinfected individuals. *Medical Journal*, 2017, Vol. 3, no. 61, pp. 95-100. (In Russ.)

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**Авторы:**

**Власова В.В.** — младший научный сотрудник лаборатории молекулярной иммунологии, Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук», г. Пермь, Россия

**Королевская Л.Б.** — к.м.н., научный сотрудник лаборатории экологической иммунологии, Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук», г. Пермь, Россия

**Логинова О.А.** — к.б.н., научный сотрудник лаборатории молекулярной иммунологии, Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук», г. Пермь, Россия

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**Authors:**

**Vlasova V.V.**, Junior Research Associate, Laboratory of Molecular Immunology, Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation

**Korolevskaya L.B.**, PhD (Medicine), Research Associate, Laboratory of Ecological Immunology, Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation

**Loginova O.A.**, PhD (Biology), Research Associate, Laboratory of Molecular Immunology, Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation

**Шмагель Н.Г.** — д.м.н., старший научный сотрудник лаборатории экологической иммунологии, Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук», г. Пермь, Россия

**Сайдакова Е.В.** — д.б.н., заведующая лабораторией молекулярной иммунологии, Институт экологии и генетики микроорганизмов Уральского отделения Российской академии наук — филиал ФГБУН «Пермский федеральный исследовательский центр Уральского отделения Российской академии наук», г. Пермь, Россия

**Shmagel N.G.**, PhD, MD (Medicine), Senior Research Associate, Laboratory of Ecological Immunology, Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation

**Saidakova E.V.**, PhD, MD (Biology), Head, Laboratory of Molecular Immunology, Institute of Ecology and Genetics of Microorganisms, Perm Federal Research Center, Ural Branch, Russian Academy of Sciences, Perm, Russian Federation

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